

Identify the Effects of Ultra Weak Light on Alphacoronavirus and Vero Cells

Hee-Chun Chung^{1,*}, Van Giap Nguyen², Hai-Quynh Do¹, Mi-Jung Park³, Jeong Su Yang³, Bong-Kyun Park¹, Woo-Kyung Jung⁴, Yong Ho Park⁴

¹Department of Veterinary Medicine Virology Lab, College of Veterinary Medicine and Research Institute for Veterinary Science, Seoul National University, Seoul, Republic of Korea.

²Department of Veterinary Microbiology and Infectious Diseases, Faculty of Veterinary Medicine, Vietnam National University of Agriculture, Hanoi, Vietnam.

³Biolight corporation in Research Center, Seoul, Republic of Korea.

⁴Noah Biotech Company in Research Center, Suwon, Republic of Korea.

Corresponding author:

H. C. Chung, Department of Veterinary Medicine Virology Lab, College of Veterinary Medicine and Research Institute for Veterinary Science, Seoul National University GwanAk-Ro 1, GwanAk-Gu, Seoul 151-742, Korea. Tel.: +82-2-880-1255, Fax: +82-2-885-0263

Keywords:

Coronavirus, Vero cells, alphacoronavirus, ultra weak light

Received: Aug 18, 2021

Accepted: Aug 19, 2021

Published: Aug 25, 2021

Editor:

Raul Isea, Fundación Instituto de Estudios Avanzados - IDEA, Hoyo de la Puerta, Baruta

Abstract

Coronavirus is causing many diseases and economic pain to humans and animals. The present study demonstrated how ultra weak light cause changes in porcine epidemic diarrhea virus (PEDV) and Vero cells.

Ultra weak light activates Vero cells by lowering pH of maintain media, but have proven no effect of killing the virus.

Introduction

Coronavirus is not only a problem for people like COVID-19, it causes a range of diseases in farm animals, some of which can be serious and are a threat to the farming industry [3]. Economically significant coronaviruses of farm animals include porcine epidemic diarrhea virus (PEDV) and bovine coronavirus (BCoV), which both result in diarrhea in young age groups animals [4]. PEDV has been circulating in Asia for several decades. It was

identified for the first time during 1980s in Japan and China [8]. PEDV became a global issue, especially in 2013, with a high death rate in the USA [9] Many researchers are working hard to get rid of PEDV, which has been a problem up until now [10].

In this study, the change was observed as a result of exposure of ultra weak light to cells and viruses respectively.

Methods

Ultra Weak Light

An optical apparatus (Photonix[®] manufactured by Biolight corporation) that emits light of ultra weak intensity was used, and its irradiance and optical spectrum were adjusted by passive optical components.

The measured irradiance at a position of 2 cm away from the center of the light emitting surface of the apparatus was about 5.2×10^{-9} W/cm² and the dominant wavelength of the spectrum was 588 nm.

Virus (PEDV)

The PEDV strain DR-13 is a PEDV strain attenuated through serial passages resulting in a 51-nucleotide deletion in the ORF3 gene [7], and it is being used for a PED oral vaccine (DR13 $10^{6.5}$ TCID₅₀/mL) commercialised in 2003 by Green Cross Vet. Prod. (South Korea).

PEDV Titration

Virus titration was carried out using a 48-well microplate with Vero cells. Virus cultures were serially diluted 10-fold with the virus replication medium containing trypsin. Confluent Vero cells of the microplate were washed three times with PBS and inoculated at 0.1 mL per well into five wells. Following adsorption for 1 h at 37 °C, the inocula were removed, and the cells were washed three times with PBS. Subsequently, 0.1 mL of fresh virus replication medium containing trypsin was transferred into each well, and the cells were further incubated for 5 days at 37 °C. Fifty per cent tissue culture infective doses (TCID₅₀) were expressed as the reciprocal

of the highest virus dilution showing cytopathic effect (CPE) [6] Antiviral effect of ultra weak light was expressed by inhibition titer, which was

calculated as follows: $[\log_{10}(\text{TCID}_{50}/\text{ml of control group}) - \log_{10}(\text{TCID}_{50}/\text{ml of experimental group})]$ [1]

Ultra Weak Light Exposure to Light with Changes in Viral Titer

Set up an experimental group that exposed 10 ml of a pre-calculated ($10^{6.5}$ TCID₅₀/ml titer) PEDV to ultra weak light and that has not been exposed to light (control group) within the CO₂ 5% incubator. Light exposure times are 10, 24, 48, 72 and 96 hours, respectively.

Test the Influence of Vero Cells on Ultra Weak Light

After incubation of approximately 1,000 Vero cells in 25T flask in the CO₂ 5% incubator, the change in the number and pH of the cells is measured in the light treated experimental group and the non-lighted control group. Light exposure times are 10, 24, 48, 72 and 96 hours, respectively.

Identification of the Inhibition of the PEDV in Vero Cells in Response to Ultra Weak Light

Incubate the 48 well plate Vero cells a day earlier and prepare an experimental group which exposed times (10, 24, 48, 72 and 96 hours) of ultra weak light and an unexposed control group.

The ultra weak light exposed experimental group after inoculation with the appropriate pre-calculated virus (titer; $10^{5.3}$ TCID₅₀/ml) was compared with the non-exposed control group.

Validation of Significance Through Statistical Analysis

All experiments were repeated 3 times. Statistical comparisons were performed using ANOVA tests ($p < 0.05$) within the SPSS program (version 15.0.0) (SPSS Inc., USA). An asterisk (*) denotes statistical significance among the mean data.

Results and discussion

Testing the Influence of PEDV on the Ultra Weak Light

We demonstrated changes in PEDV titers during exposure times for ultra weak light.

Following exposure to ultra weak light, the PEDV inhibition above the logarithmic reduction = $\log 0.1$ (20.567%) from 10h to 72h was confirmed, but the statistical analysis was not significant (Figure 1). The effectiveness of disinfectant as a disinfectant announced a maximum reduced titer, in which the virus titer was reduced by at least 4 \log_{10} [2,5] was not met to be an effective dilution factor.

Testing the Influence of Vero Cells in Response to the Ultra Weak Light

We examined how Vero cells changes following exposure to ultra weak light.

After exposure to ultra weak light, the pH level maintained lower than the control group for 10 to 96 h, which enabled Vero cells, increasing the number of cells by 1.2 to 1.5 times more than the control group. In particular, differences due to larger effects were identified during light exposure for over 72 hours, indicating that the statistical analysis was also significant (Figure 2). The ultra weak light activated the cell by decreasing the pH of the growth medium.

Identification of PEDV Inhibition in Vero Cells in Reaction to Ultra Weak Light

We found changes in PEDV in Vero cells exposed to ultra weak light.

After exposure to ultra weak light, the virus inhibition of \log reduction=0.1 (20.567%) from 10h to 48h was confirmed, but statistical analysis showed no significant effects of virus reduction (Figure 3).

Funding

This work was supported by the Korea Science and Engineering Foundation (KOSEF) grant funded by the Korea government (No. 2020R1I1A1A01054539) and by Korea Institute of Planning and Evaluation for Technology in Food, Agriculture and Forestry (IPET) through Technology Commercialization Support Program funded by Ministry of Agriculture, Food and Rural Affairs (MAFRA) (No. 821065-03), respectively.

Acknowledgments

The authors would like to thank Eun Ok Kim and Jung Ah Kim for their excellent technical assistance.

Conflicts of Interest

The authors declare that there is no conflict of interest.

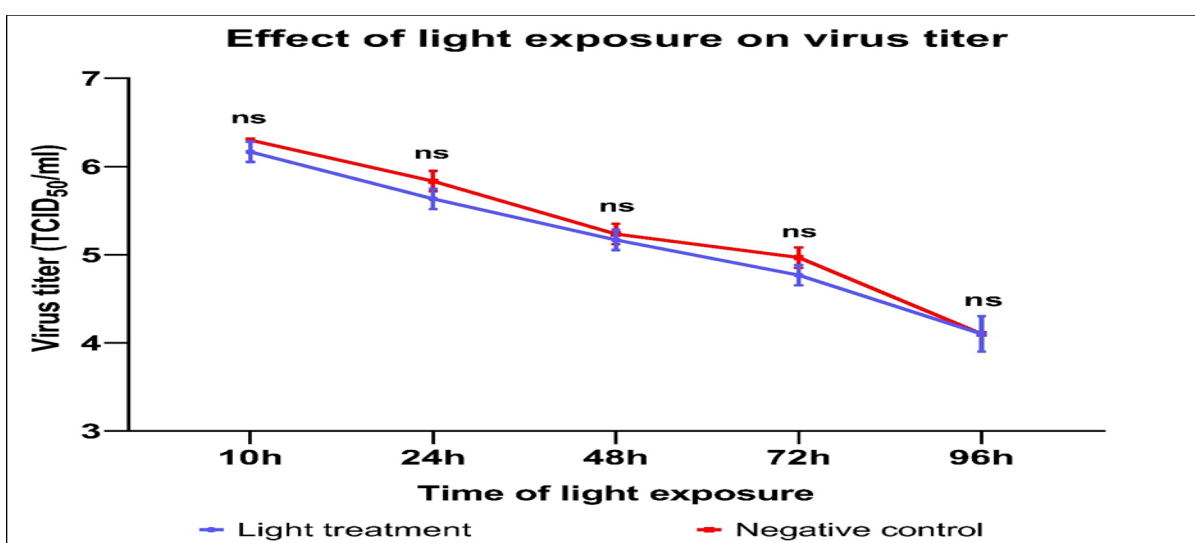


Figure 1. Changes in PEDV titers according to ultra weak light exposure times
ns mean: non-significant

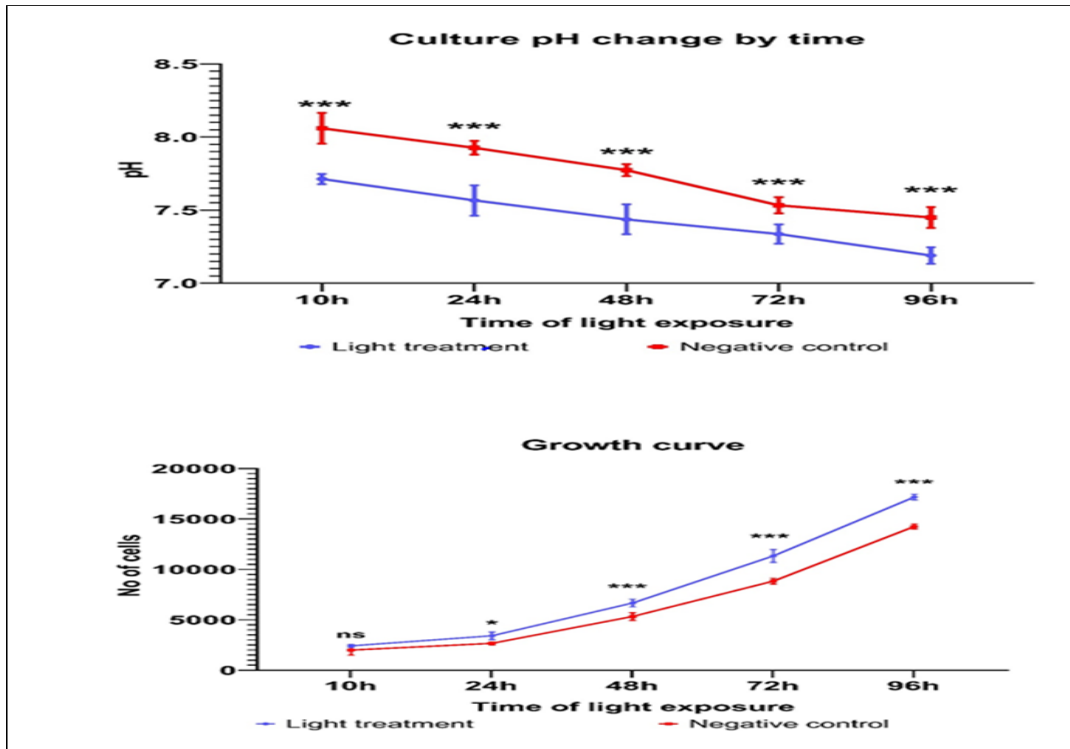


Figure 2. Variation of Vero cell and pH due to exposure to ultra weak light
 ns mean : non-significant, * mean significant at p-value < 0.05, *** mean significant at p-value < 0.005

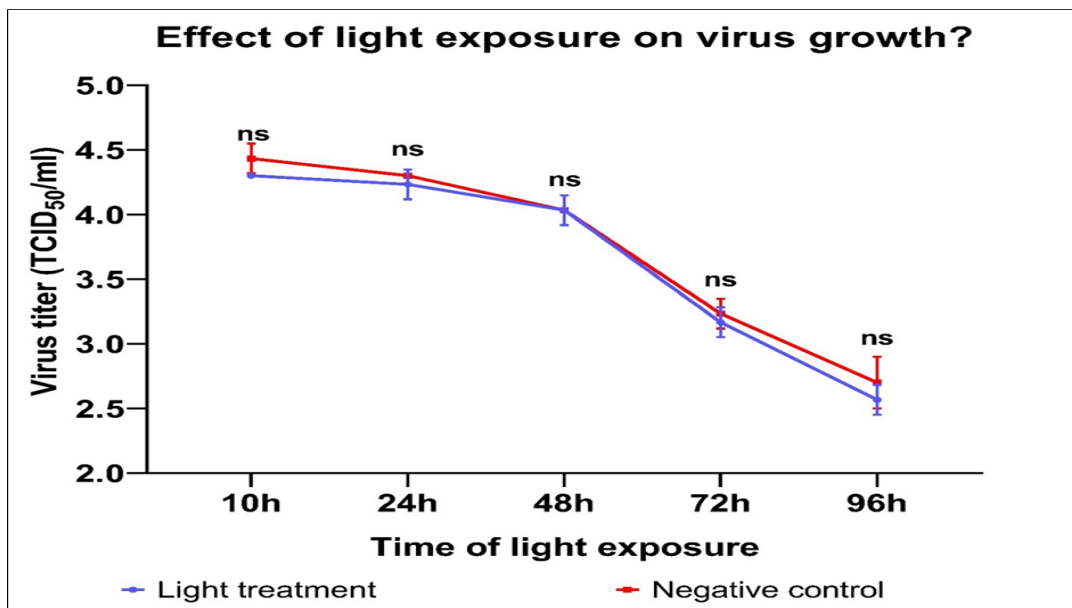


Figure 3. PEDV Inhibition in exposed ultra weak light within Vero Cells

References

1. Chen, Y.-N., Hsueh, Y.-H., Hsieh, C.-T., Tzou, D.-Y., Chang, P.-L., 2016. Antiviral activity of graphene-silver nanocomposites against non-enveloped and enveloped viruses. *International journal of environmental research and public health* 13, 430.
2. de Oliveira, T.M.L., Rehfeld, I.S., Guedes, M.I.M.C., Ferreira, J.M.S., Kroon, E.G., Lobato, Z.I.P., 2011. Susceptibility of Vaccinia virus to chemical disinfectants. *The American journal of tropical medicine and hygiene* 85, 152.
3. Esudu, E.L., 2021. A survey on the impact of Covid19 on Animal production and marketing. Busitema University.
4. Haake, C., Cook, S., Pusterla, N., Murphy, B., 2020. Coronavirus infections in companion animals: virology, epidemiology, clinical and pathologic features. *Viruses* 12, 1023.
5. Jimenez, L., Chiang, M., 2006. Virucidal activity of a quaternary ammonium compound disinfectant against feline calicivirus: a surrogate for norovirus. *American journal of infection control* 34, 269-273.
6. KUSANAGI, K.-i., KUWAHARA, H., KATOH, T., NUNOYA, T., ISHIKAWA, Y., SAMEJIMA, T., TAJIMA, M., 1992. Isolation and serial propagation of porcine epidemic diarrhea virus in cell cultures and partial characterization of the isolate. *Journal of Veterinary Medical Science* 54, 313-318.
7. Park, S.-J., Moon, H.-J., Luo, Y., Kim, H.-K., Kim, E.-M., Yang, J.-S., Song, D.-S., Kang, B.-K., Lee, C.-S., Park, B.-K., 2008. Cloning and further sequence analysis of the ORF3 gene of wild-and attenuated-type porcine epidemic diarrhea viruses. *Virus genes* 36, 95-104.
8. Song, D., Park, B., 2012. Porcine epidemic diarrhoea virus: a comprehensive review of molecular epidemiology, diagnosis, and vaccines. *Virus genes* 44, 167-175.
9. Vlasova, A.N., Marthaler, D., Wang, Q., Culhane, M.R., Rossow, K.D., Rovira, A., Collins, J., Saif, L.J., 2014. Distinct characteristics and complex evolution of PEDV strains, North America, May 2013–February 2014. *Emerging infectious diseases* 20, 1620.
10. Yang, J.-L., Ha, T.K.Q., Oh, W.K., 2016. Discovery of inhibitory materials against PEDV corona virus from medicinal plants. *Japanese Journal of Veterinary Research* 64, S53-S63.