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Research Article

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Morphological Features of Testicular Biopsies in Infertile Males at a Tertiary Hospital, Southwest Nigeria

ADESOJI ADETONA^{1,*}, Olufemi Ogunbiyi¹

¹Nigeria

Corresponding author:

ADESOJI ADETONA, Nigeria

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Amit Kant, Department of Physiology, U.P. University of Medical Sciences, House No. 168, Kaveri Kunj, Phase II, Kamla Nagar, Saifai, Etawah, Agra, 282005, Uttar Pradesh, India.

Abstract

Background

Male Infertility accounts for 30-40% of all cases of infertility and its evaluation requires a good history, thorough physical examination, and several investigations to include testicular biopsy which might be used to further categorize infertile males for proper management and prognostication. This study aims to determine the predominant histopathological patterns of testicular biopsies in infertile males and to compare the findings with previous studies.

Methods

A retrospective cross-sectional study of 225 selected cases of testicular biopsies reviewed for the evaluation of male infertility in the Pathology department, of a tertiary hospital, Southwest, Nigeria, between 1987 and 2012. Relevant clinical and histopathological information was extracted from the departmental records. All histologic cases were reviewed, and a classification based on histological patterns of spermatogenesis was utilized to group the cases into normal findings, hypo spermatogenesis, maturation arrest, Sertoli cell-only syndrome, peritubular hyalinization/ tubular fibrosis and mixed patterns. The data obtained were analysed using descriptive and inferential statistics at a 5% level of significance.

Results

Among the 225 cases reviewed with a mean age of 37.7 years (SD - 8.61), 82.7% had primary infertility of which 92.9% were azoospermic, while 7.1% had oligospermia. The histological patterns included 34.2% of Hypospermatogenesis, 32% of Peritubular hyalinization/ tubular fibrosis, 14.2%





had maturation arrest and Sertoli cell-only syndrome was found in 6.7% of cases, only 0.9% had normal histologic pattern while the mixed histologic pattern was seen in 12% of cases.

Conclusion

The commonest morphological pattern was Hypospermatogenesis, which is similar to some of the previous local and international studies. A high percentage of peritubular fibrosis was noted with few tubules containing scanty late spermatids or spermatozoa when proper sampling and evaluation were made. Multiple patterns within a biopsy were seen with careful review, especially in non-obstructive azoospermic cases. This is significant in male infertility patient management in our environment because it suggests greater chances of successful sperm extraction for Assisted Reproduction Technique in such patients.

Background

Infertility is a medical and socio-cultural phenomenon that affects up to 15% of couples worldwide with male factors accounting for 30-40% of all cases of infertility.[1] Its evaluation requires a comprehensive history and thorough physical examination along with investigations such as semen analysis, hormonal assay and testicular biopsy.[2,3]

The causes of male infertility can be grouped into three categories: Pre-testicular, Testicular and Post-testicular causes.

The pre-testicular causes include extragonadal endocrine disorders originating in the hypothalamus, pituitary or adrenal glands; chronic illnesses, and certain medications such as some antihypertensive drugs like Nifedipine as well as antiviral agents like Ganciclovir and a few others.[1,4]

The testicular causes are related to defects in the process of spermatogenesis such as maturation arrest, germ cell aplasia, etc.[4]

The post-testicular causes include obstruction of ducts

draining the testes secondary to trauma or surgery, impaired sperm motility, morphological abnormalities of the spermatozoa and biochemical abnormality of the seminal fluid.[1,5]

Biopsy Technique

The FNA procedure is performed on the testis by aspiration at three separate sites (upper, middle and lower pole). Four to five needle excursions were made at each site except when contraindicated[6]. Contraindications for bilateral testicular sampling included the presence of local skin infection, hydrocele, orchialgia or a previous biopsy[6,7]. Another technique is by open biopsy at the same site of the needle biopsy. Both needle specimen and open tissue specimen are fixed in Bouin's solution and sent for histologic examination. Routine haematoxylin and eosin (H&E) for the open tissue biopsy and Romanowsky stains is performed on the smears[8].

An open testicular biopsy shows different morphological patterns, which can provide valuable information to the urologist to further categorize infertile males with azoospermia or oligospermia for prognostication and treatment. Whereas, obstructive and testicular causes usually show a narrow range of morphologic changes, which could be assessed with biopsy evaluation to determine the patient reproductive chances and treatment options.

This study aimed to determine the predominant histopathological patterns seen in the testicular biopsies of infertile males and to compare the findings with those seen in similar studies including a previous study in this centre.

This information might be useful in the management of cases of male infertility by guiding case selection of patients that can benefit from in vitro fertilization or surgical intervention in cases where especially there is duct obstruction with the normal histologic pattern.





Methods

This is a retrospective cross-sectional study carried out in the Department of Pathology of a large teaching hospital in South-Western Nigeria. We reviewed all testicular biopsies submitted for the evaluation of male infertility between 1987 and 2012 received from both the urology clinic of the hospital and peripheral hospitals, usually fixed either in Bouin's fluid or 10% buffered formalin. The archival Haematoxylin & Eosin (H&E) stained histology slides of cases were retrieved. Cases whose slides and blocks were missing were excluded from the study. All the cases of fibrosis were easily diagnosed on H&E and there was no need for any special stains such as Periodic Acid Schiff or Verhoeff van Gieson. All histologic cases were reviewed and a morphological classification that is principally based on histological patterns of spermatogenesis was utilized to group the cases into normal findings, hypo spermatogenesis, maturation arrest, Sertoli cell-only syndrome, peritubular hyalinization/ tubular fibrosis and mixed patterns[2,9,10]. The data obtained were analysed using descriptive and inferential statistics at a 5% level of significance and Ethical approval was obtained from the appropriate Ethical Review Committee.

Results

A total of 225 cases were reviewed comprising of patients who presented with infertility and had either azoospermia or oligospermia diagnosed on routine seminal fluid analysis.

Their ages ranged from 20 years to 60 years with a mean age of 37.7 years (SD = 8.61). The highest proportion of the cases, 31.6%, was found among the age group 30-39 years. The majority, (82.7%), had primary infertility while 17.3% had secondary infertility.

Seminal fluid analysis reports had shown that most of the cases reviewed had azoospermia, (92.9%), while others had oligospermia. Only 9.8% had clinical evidence of obstruction. Azoospermia was seen in 93% of the primary and 92.5% of the secondary infertility cases. All the cases were histologically classified into six categories (figure 1).

Hypospermatogenesis (Hypo) was the most common pattern seen comprising 34.2%, followed by peritubular hyalinization/ tubular fibrosis (PHTF) in 32%, maturation arrest (MA) in 14.2%, Sertoli cell-only syndrome (SCOS) in 6.7%, while the least pattern were cases of normal histologic pattern in 0.9%. The photomicrographs of each pattern are shown in figure 2 below. Twenty-seven cases (12%) had a mixed histological pattern where more than one morphologic picture was seen in the same biopsy (Figure 3).

In relating the different histologic patterns with the types of infertility, more cases of primary infertility were seen in all the different histologic patterns compared with secondary infertility. Peritubular hyalinization/tubular fibrosis is the commonest pattern seen among cases of primary infertility (87.5%) while Hypospermatogenesis is the commonest in the secondary type (43.6%). The two histologically normal cases also had primary infertility (100%). Clinical evidence of obstruction was seen to be more associated with hypo spermatogenesis, (40.9%) and maturation arrest, (31.8%) (Table 1).

In this study, the mixed histologic pattern constituted 12% of the entire cases reviewed (Table 2) with various combinations which include peritubular hyalinization/tubular fibrosis with hypo spermatogenesis and peritubular hyalinization/tubular fibrosis with maturation arrest in 29.6% each, peritubular hyalinization/tubular fibrosis with Sertoli cell-only syndrome in 18.6%, both hypo spermatogenesis with Sertoli cell-only syndrome and maturation arrest with Sertoli cell-only syndrome in 11.1% of cases each.

Most of the mixed histologic pattern cases, 77.8%, had primary infertility. Table 3 showed the association between the mixed histologic patterns, semen analysis report and evidence of obstruction. However, these relationships are not statistically significant ($p \ge$ 0.05)





Table 1. Correlation between Histologic Patterns, SFA, and Evidence of Obstruction									
Histologic Patterns	Type of Infertility			SFA			Obstruction		
	Primary	Secondary	Total	Azoospermia	Oligo spermia	Total	No	Yes	Total
PHTF	64(28.4%)	8(3.6%)	72(32.0%)	67(29.8%)	5(2.2%)	72(32.0%)	69(30.7%)	3(1.3%)	72(32.0%)
НҮР	60(26.7%)	17(7.6%)	77(34.2%)	72(32.0%)	5(2.2%)	77(34.2%)	68(30.2%)	9(4.0%)	77(34.2%)
MA	25(11.1%)	7(3.1%)	32(14.2%)	28(12.4%)	4(1.8%)	32(14.2%)	25(11.1%)	7(3.1%)	32(14.2%)
MIXED	22(9.8%)	5(2.2%)	27(12.0%)	25(11.1%)	2(0.9%)	27(12.0%)	24(10.7%)	3(1.3%)	27(12.0%)
SCOS	13(5.8%)	2(0.9%)	15(6.7%)	15(6.7%)	0(0.0%)	15(6.7%)	15(6.7%)	0(0.0%)	15(6.7%)
Normal	2(0.9%)	0(0.0%)	2(0.9%)	2(0.9%)	0(0.0%)	2(0.9%)	2(0.9%)	0(0.0%)	2(0.9%)
Total	186(82.7%)	39(17.3%)	225 (100.0%)	209(92.9%)	16(7.1%)	225 (100.0%)	203(90.2%)	22(9.8%)	225 (100.0%)

Table 1. Correlation between Histologic Patterns, SFA, and Evidence of Obstruction

Table 2. Frequency of Mix	xed Pattern	
MIXED PATTERNS	FREQUENCY	PERCENTAGE
PHTF+HYPO	8	29.6
PHTF+MA	8	29.6
PHTF+SCOS	5	18.6
SCOS+HYPO	3	11.1
SCOS+MA	3	11.1
Total	27	100.0



Table 3. Correlation between Mixed Histologic Patterns, SFA, and Evidence of Obstruction

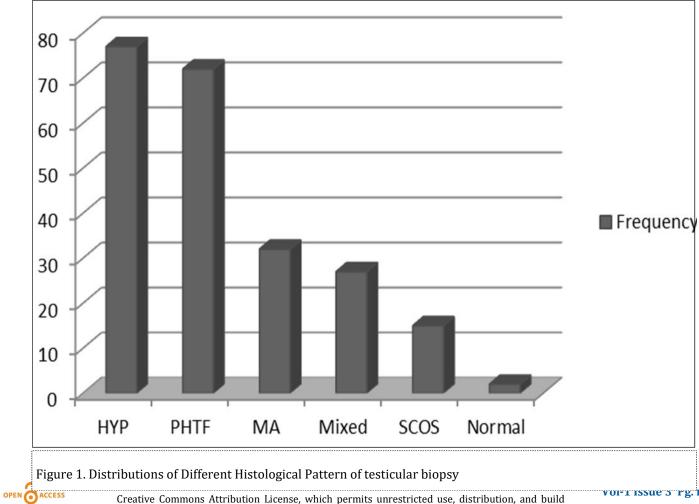
Type of		fertility		SFA			Obstruction			
Histologic Patterns	Primary	Secondary	Total	Azoo- spermia	Oligo- spermia	Total	No	Yes	Total	
Single Pattern	165 (72.9%)	34 (15.1%)	198 (88.0%)	184 (81.8%)	14 (6.2%)	198 (88.0%)	179 (79.6%)	19 (8.4%)	198 (88.0%)	
HYP+SCO S	6 (2.7%)	2 (0.9%)	8 (3.6%)	3 (1.3%)	0 (0.0%)	3 (1.3%)	3(1.3%)	0(0.0%)	3 (1.3%)	
MA+SCOS	6 (2.7%)	2 (0.9%)	8 (3.6%)	2 (0.9%)	0 (0.0%)	2 (0.9%)	2(0.9%)	0(0.0%)	2 (0.9%)	
SCOS+PH TF	5 (2.2%)	0 (0.0%)	5 (2.2%)	4 (1.8%)	1 (0.4%)	5 (2.2%)	4(1.8%)	1(0.4%)	5 (2.2%)	
MA+ PHTF	2 (0.9%)	1 (0.4%)	3 (1.3%)	7 (3.1%)	1 (0.4%)	8 (3.6%)	8(3.6%)	0(0.0%)	8 (3.6%)	
HYP+ PHTF	1 (0.4%)	0 (0.0%)	1 (0.4%)	9 (4.0%)	0 (0.0%)	9 (4.0%)	7(3.1%)	2(0.9%)	9 (4.0%)	
Total	186 (82.7%)	39 (17.3%)	225 (100.0%)	209 (92.9%)	16 (7.1%)	225 (100.0%)	203 (90.2%)	22 (9.8%)	225 (100.0%)	





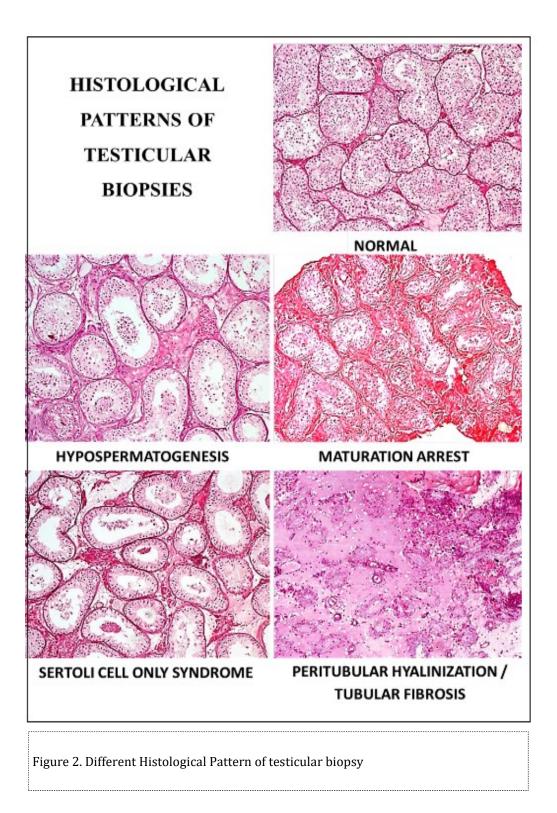
	Nor- mal	Нуро	MA	SCOS	PHTF	Mixed	Klinefel-ter synd.	Inflam- mato- ry	Varico-coele
Present Study(n=225)	0.9	34.2	14.2	6.7	32	12	-	-	-
Thomas Ibadan (n=152)	38.2	19.1	5.3	9.2	23	-	5.3	-	-
Obafunwa et al. Lagos (n=177)	20	49	-	1.1	-	-	1.1	24.3	3
Ahmed et al. Za- ria (n=472)	15.9	35.2	14.6	28.2	6.1	-	-	-	-
Ojo et al. Ilorin (n=63)	14.1	4.9	25.4	-	55.6	-	-	-	-
Ali et al. Egypt (n=272)	5	39	1.0	26	19	-	-	-	-
Al-Rayess et al. Saudi(n=230)	31.4	13	11	39.3	5.3	-	-	-	-
Nagpal et al. In- dia	16	42	18	17	1	-	3	3	-

Table 4. Geographical Variation of Histologic Patterns in male infertility



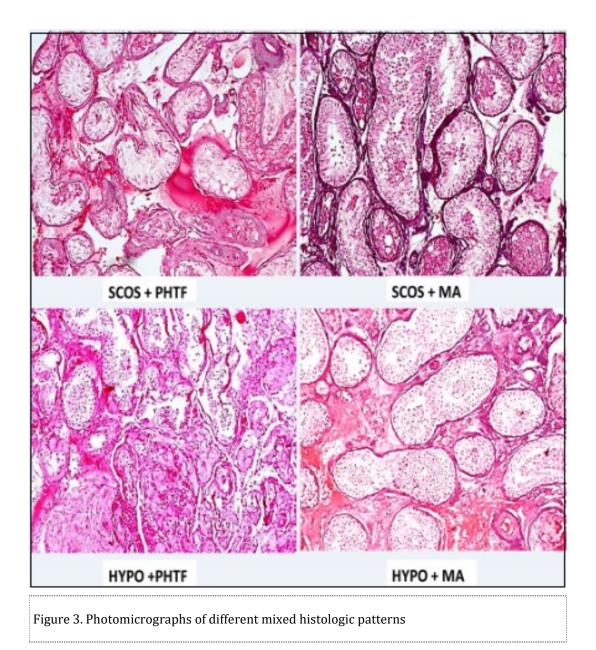
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Discussion

We reviewed testicular biopsy specimens from 225 men diagnosed by seminal fluid analysis to be either azoospermic or oligospermic. The mean age of the cases was 37.7 years (SD 8.61, range 20-60 years[11,12] Primary infertility was commoner than secondary infertility in this study, as was the pattern seen in a few previous studies elsewhere in Nigeria.[13,14] reported 49.2% of patients with primary infertility and 22.2% with secondary infertility in llorin. Couples with primary infertility tend to present early for fertility evaluation in the health centre due to socio-cultural factors and family pressures than those with secondary infertility who had once had a pregnancy to term irrespective of the outcome or those with one or more children in this environment[13].

As shown, the majority of cases were in the age range 30-39 years corresponding to the peak of male reproductive activity and the age of psychological maturity when infertile men are most concerned about their condition and seek medical attention[15].

In this study, 209 (92.9%) patients were azoospermic by the SFA report while others were oligospermic. Also, the clinical history of obstruction and clinical evidence of obstruction was noted in 10% of cases with azoospermia while none of the oligospermic patients had any associated clinical obstruction. This finding is similar to what was reported in some of the previous local and international studies.[16,17] The evaluation of the possible aetiopathogenesis of testicular damage in non-obstructive azoospermia, which has a high incidence in this environment may help in the prevention and management of male infertility.

Normal spermatogenesis has been seen in a variable proportion of testicular biopsies done in the investigation of primary infertility. These usually represent post-testicular problems in such patients (including obstruction). In many studies, these were reported in up to between 33% and 50% of cases.

[18-20] The frequency of normal spermatogenesis in the present study is much lower at 0.9%.

Hypospermatogenesis (Hypo) was the most common pattern seen in this study constituting 34.2% of the cases. A similar pattern but with a higher proportion was seen in the study by Obafunwa et al. from Lagos [5], Ahmed et al from Zaria[11] and Nagpal et al. from India [21] while a lower prevalence was seen in other previous studies done in this environment including the previous study from this centre by Thomas[18] who reported 19% of cases. Ikuerowo et al[22] from Lagos, Ojo et al[14] from Ilorin and Oranusi et al[23] from Nnewi reported 3.9%, 4.8% and 10% of cases with Hypospermatogenesis respectively. Various reasons were adduced for the sharp differences in the percentages by some of these authors, which are also similar to the reasons identified among the few that are specified in this study too, which include exposure to noxious chemicals (lead, various toxic industrial fumes especially gasoline vapour), drugs e.g. Nifedipine, Moduretic and physical agents (especially heat, tightly fitted underwear which apposes the testes closely to the body).[3,5,13]

Maturation arrest (MA) was observed in 32 (14.2%) cases with an arrest at different levels of maturation, none reaching up to the level of spermatid. Those reviewed in this study were at the level of spermatogonia and primary spermatocytes. The incidence of maturation arrest was similarly lower in the previous study from Ibadan where 5.3% was reported. Philip et al[12] reported 5.9% and a very low incidence of 0.4% was reported by Ali from Saudi.[24] However, a higher relative proportion of 27.5% and 25.4% were reported by Ikuerowo et al[22]and Ojo et al. from Lagos and Ilorin respectively.[14].

Sertoli cell-only syndrome (SCOS) was found in 6.7% of cases. This pattern usually has a very low incidence and was so documented by Thomas[18] in Ibadan who reported 9.2% and by Obafunwa et al[5] and Phillip et al[12] who documented 1.1% and 1.5%



respectively. In contrast, some of the studies reported higher incidences. Ahmed et al[11] from Zaria reported 28.2% of cases, Rasheed et al.[4] from Egypt reported 34% of cases. In India, Mehrotra et al[19] and Nagpal et al [21] reported 29.3% and 17% respectively.

Peritubular hyalinization/tubular fibrosis (PHTF) was found in 32% of the cases in this study. This was less frequent than in the previous study done by Thomas with 23% cases.[18]. Two recent studies from Lagos by Ikuerowo et al[22] and Ojo et al[14] similarly reported high incidences of 58.8% and 55.6% while some other studies reported low incidences such as those of Ahmed et al[11] from Zaria (6.1%) and Rasheed et al[4] from Egypt (6.0%). Al-Rayess et al[16] and Ali et al[24] from Saudi reported 12% and 19% respectively.

A mixed morphological pattern was diagnosed in this study when over 30% of either pattern was present and this was the case in 28 (12%) patients. Of these, there were 9 cases of peritubular hyalinization/tubular fibrosis admixed with hypo spermatogenesis; 8 cases of peritubular hyalinization/tubular fibrosis mixed with Maturation arrest; 5 cases of peritubular hyalinization/ tubular fibrosis mixed with Sertoli cell-only syndrome; 3 cases of Sertoli cell-only syndrome mixed with hypo spermatogenesis and 3 cases of Sertoli cell-only syndrome mixed with maturation arrest.

The limitation of this study includes the smaller number of samples over a long period, absence of fertility hormone profile and lack of molecular or genetic testing capability of the institution to further screen the participants for genetic anomalies for a better correlation with morphologic patterns seen.

Some studies identified different discordant patterns in right and left testes, supporting the use of bilateral testicular biopsies for a more comprehensive evaluation of male infertility. Although the absence of spermatozoa in one testis in such cases does not rule out their presence in the other[2] a unilateral biopsy has been seen to suffice in revealing the pathology in most instances[2,25]. A bilateral biopsy seems needful only when there is a significant difference in the size of the testes and especially when considering sperm retrieval for an intracytoplasmic sperm injection (ICSI) in the management of infertile couples with male infertility[26-28].

From the literature, there is significant geographical variation in the histological patterns in male infertility as shown in Table 4

The high prevalence of normal spermatogenesis in a previous study was attributed to the high incidence of chronic inflammation resulting in fibrosis of the excurrent ducts. This seems to occur with much lower frequency in recent studies due to the advent of indiscriminate antibiotic use in our environment which has reduced the incidence of chronic inflammatory patterns seen in these patients.[5,21] Besides, Klinefelter syndrome is reported less frequently in recent studies due to improved methods for early diagnosis including in-utero diagnosis negating the need for testicular biopsy in the evaluation of such patients for infertility.[5,18,21]. In another study, 2.7% had chromosomal abnormalities, of which less than half had sex chromosome abnormalities (mainly 47, XXY) while Y-chromosome microdeletions were detected in 5.4%.[29]

The variations observed between this study and some other previous studies are not well understood. However, previous local and international studies have suggested increasingly, the importance of occupation, environmental, sociocultural habits, consanguineous marriages, and in particular, genetic factors in the cause of male infertility[30,31,32].

Proper evaluation of the testicular biopsy of infertile males is very necessary for the management of such patients and pathologists are required to report any form of germ cells seen in the specimen, especially if late spermatids or spermatozoa are seen. Quantitative assessment of human spermatogenesis has been shown by recent studies to be possible and can be beneficial.[33] This is very important in offering the chance of assisted



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fertilization that will require testicular sperm extraction and intracytoplasmic sperm injection, and also for some patients with smaller testicles and slightly elevated levels of FSH.[34]

Conclusion

The findings of this study are similar to some of the previous local and international studies.

Most infertile males had primary infertility and azoospermia, and the commonest identified histologic pattern of testicular biopsy in cases of male infertility identified in this study was hypo spermatogenesis, followed by peritubular hyalinization/tubular fibrosis.

This study has shed some light on the possibility of multiple patterns within a biopsy if properly sampled and reviewed carefully especially in non-obstructive azoospermic cases with greater chances of successful sperm extraction for Assisted Reproduction Technique.

Bilateral testicular biopsies, as well as meticulous pathological examination of all seminiferous tubules in view, should be carried out to identify mixed and discordant patterns and this is highly recommended.

This study was self-funded with no support from any quarters.

List of Abbreviations

H&E - Haematoxylin & Eosin

- FSH Follicle Stimulating Hormone
- SFA Seminal Fluid Analysis
- HYP-Hypospermatogenesis

PHTF–Peritubular Hyalinization/ Tubular fibrosis MA–MaturationArrest

Mixed-MixedPattern

SCOS–Sertoli Cell Only Syndrome

Normal – Normal Pattern

ICSI - Intracytoplasmic Sperm Injection.

References

- 1. Cerilli LA, Kuang W RD, Cerilli LA, Kuang W, Rogers D.
 - A practical approach to testicular biopsy

interpretation for male infertility. *Arch Pathol Lab Med*. 2010;134(8):1197–204.

- Abdullah L, Bondagji N. Histopathological patterns of testicular biopsy in male infertility: A retrospective study from a tertiary care center in the western part of Saudi Arabia. *Urol Ann*. 2011;3:19–23.
- Naz M, Kamal M. Classification, causes, diagnosis and treatment of male infertility: a review. Oriental Pharmacy and Experimental Medicine. 2017.
- Rashed M, Ragab N, Shalaby A RW. Patterns Of Testicular Histopathology In Men With Primary Infertility. *Internet J Urol.* 2008;5(2):1–5.
- Obafunwa JO, Elesha SO Morphological changes found in the testes of 177 Nigerian males investigated for infertility. *Afr J Med Med Sci.* 1993;22(4):35–40.
- Mehrotra R, Chaurasia D. Fine needle aspiration cytology of the testis as the first-line diagnostic modality in azoospermia: a comparative study of cytology and histology. 2008;363–8.
- Cohen MS, Frye S, Warner RS, Leiter E. Testicular needle biopsy in diagnosis of infertility. *Urology*. 1984;24:439–42.
- Craft I, Tsirigotis M, Farrer-brown G. Testicular needle aspiration as an alternative to biopsy for the assessment of spermatogenesis. 1997;12(7):1483–7.
- Ticko SK, Amin MB, Cramer HM, Harik LR, Ulbright TM. The testis, paaratesticular structures, and male external genitalia. In: Silverberg SG, DeLellis RA, Frable WJ, LiVolsi VA WM, editor. Principles and Practice of Surgical Pathology and Cytopathology. fourth. Philadelphia: Elsevier Churchill Livingstone; 2006. p. 1731–89.
- Mushtaq H, Alam S, Khan MA. Histopathological Patterns of Testicular Biopsies in Male Infertility. J Islam Med Dent Coll. 2013;
- Ahmed SA, Mohammed A, Shehu SM, Samaila MOA, Mbibu NH, *et al.*, Morphological Pattern of Testicular Biopsies in Zaria, Nigeria. *Niger Med J*. 2007;48(3):69





-70.

- Philip C, Bilodi AK PS. Histological Evaluation of Testicular Biopsies and Spermatogenesis Distubance in Infertile Men with Varicocoele. *World J Pharm Sci.* 2014;3(3):1789–803.
- Deye Vincent, F., Michel, P., Ehrmann, S., Da Silva, D., Piagnerelli, M., *et al.*, (2016). Changes in cardiac arrest patients TM temperature management after the 2013 Hussaini & Mustafa, S. (2016). Adolescentsâ€TM knowledge and awareness of diabetes mellitus in Kuwait. Alexandria Journal of Medicine, 52(1), 61–66. M,
- 14. Ojo BA, Rahman GA The Value of Testicular Biopsy in Male Infertility: Experience with 63 Nigerians. *African J Urol.* 2005;11(2):105–10.
- 15. Articles on Psychology, Psychiatry and M. Psychological Maturity [Internet]. New Horizons. Available from: http://sabryfattah.com/psychology/ psychological-maturity/
- Al-Rayess, Molham M, Al-Rikabi AC. Morphologic patterns of male infertility in Saudi patients: A University Hospital experience. *Saudi Med J*. 2000;21:625–8.
- Colgan TJ, Bedard YC, Strawbridge HT, Buckspan MB, Klotz PH. Reappraisal of the value of testicular biopsy in the investigation of infertility. *Fertil Steril*. 1980;33:56–60.
- Thomas JO. Histological pattern of testicular biopsies in infertile males in Ibadan, Nigeria. *East Afr Med J.* 1990;67(8):578–84.
- Mehrotra, R. Chaurasia D. Fine needle aspiration cytology of the testis as the first-line diagnostic modality in azoospermia: a comparative study of cytology and histology. *Cytopathology*. 2008;19 (6):363–368.
- 20. Caucci M, Barbatelli G CS. The retractile testis can be a cause of adult infertility. *Fertil ity Steril*. 1997;68 (6):1051–8.

- Nagpal BL, Manjari M, Kapoor K, Dhaliwal US. Testicular biopsy in cases of male infertility: a retrospective study. *J Indian Med Assoc.* 1993;91:171– 4.
- Ikuerowo SO, Izegbu MC, Benebo AS, Fadeyibi IO, Omodele FO. Testicular Biopsies of Azoospermic Men at The Lagos State University Teaching Hospital. *African J Urol.* 2010;16(3):69–72.
- Oranusi CK, Onyiaorah IV, Ukah CO. Pattern of Testicular Biopies as Seen in a Tertiary Institution in Nnewi, Southeast Nigeria. *Niger J Surg.* 2014;20(2): 55–8.
- Ali MA, Akhtar M, Woodhouse N, Burgess A, Faulkner C, *et al.,*. Role of testicular fine-needle aspiration biopsy in the evaluation of male infertility: cytologic and histologic correlation. *Diagn Cytopathol.* 1991;7 (2):128–31.
- 25. Brannen GE, Roth RR. Testicular abnormalities of the subfertile male. *J Urol*. 1979;122:757–62.
- McLachlan RI, Rajpert-De Meyts E, Hoei-Hansen CE, de Kretser DM, *et al*,.. Histological evaluation of the human testis—approaches to optimizing the clinical value of the assessment. *Hum Reprod.* 2007;22(1): 2–16.
- Levin HS. Testicular biopsy in the study of male infertility: its current usefulness, histologic techniques, and prospects for the future. *Hum Pathol*. 1979;10:569–84.
- Venkatachala S, Malur PR, Nerli RB, Desai BR, Dhorigol V. Testicular biopsies--histomorphologic patterns in male infertility. *Indian J Pathol Microbiol*. 2007;50:726–9.
- 29. Olesen IA, Andersson AM, Aksglaede L, Skakkebaek NE, Rajpert-de Meyts E, Joergensen N, et al. Clinical, genetic, biochemical, and testicular biopsy findings among 1,213 men evaluated for infertility. *Fertil Steril*. 2017;
- 30. Irvine, D. Stewart. Epidemiology and aetiology of





male infertility. Human Reproduction. 1998. p. 33-44.

- Dada R, Kumar M, Jesudasan R, Fernández JL, Gosálvez J, *et al.,co* Epigenetics and its role in male infertility. Journal of Assisted Reproduction and Genetics. 2012. p. 213–23.
- Krausz C, Escamilla AR, Chianese C. Genetics of male infertility: From research to clinic. Reproduction. 2015.
- Jezek D, Banek L, Hittmair A, Pezerović-Panijan R, *et al.,*. Mast cells in testicular biopsies of infertile men with "mixed atrophy" of seminiferous tubules. *Andrologia*. 1999;31(4):203–10.
- 34. Corradi G, Kiss F, Borka K, Füle B, Szabó K. Current viewpoints on the evaluation of testicular biopsy. Orv Hetil. 1998;139:2243–6.

